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Highly regioselective ring opening of epoxides using NaN₃: a short and efficient synthesis of (–)-cytoxazone

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Abstract—A convenient and efficient synthesis of 1,2-azido alcohols has been achieved by regioselective ring opening of epoxides using NaN₃ and 4Å molecular sieves in acetonitrile. The utility of this method has been demonstrated by achieving a short synthesis of (-)-cytoxazone in 48% overall yield.

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1,2-Azidoalcohols are versatile intermediates in organic synthesis, since they are important precursors of β -aminoalcohols and vicinal diamines, which are present in numerous natural products.¹ Furthermore, their importance is significant in the chemistry of carbohydrates and nucleosides.² Since the discovery of Sharpless asymmetric epoxidation³ and asymmetric dihydroxylation⁴ the ring opening reactions of epoxides have been extensively studied in the context of the chiral synthesis of biologically active natural products.

Generally, azidohydrins are prepared through ring opening of epoxides using different azides in suitable solvents. The classical method uses⁵ sodium azide and ammonium chloride but it requires a long reaction time (12–48 h) and is often accompanied by isomerization, epimerization and rearrangement of the products.^{6a} Hence, several modifications have been made, which include the combined use of NaN₃ or TMSN₃ and a Lewis acid or transition metal complex⁷ or reaction in the presence of tetrabutylammonium salts.⁸ Recently, pH-controlled azidolysis of epoxides was also studied in water using various Lewis acids and NaN₃.⁶

Although these methods have their own advantages, many of them suffer from one or more drawbacks such

as long reaction times, strongly acidic conditions, lack of regioselectivity, high cost of reagent, difficulty in workup and lack of reusability of a catalyst. Therefore a better catalyst that would be superior to the existing ones, especially with regard to recyclability and selectivity, is still required.

In recent years, organic reactions on solid supports such as zeolites or mesoporous molecular sieves⁹ have attracted attention because of the advantages these catalysts have, such as high purity of the product, easy work-up procedure, reusability and the environmentfriendly nature of the catalyst. In continuation of our interest in the chemistry of epoxides and their application in the synthesis of bioactive natural products,¹⁰ we found that a variety of epoxides can be opened in a regioselective manner with NaN₃ using 4Å molecular sieves as the catalyst (Table 1).

It is evident from Table 1 that in the case of terminal epoxides such as 1, 2, 4, 5 and 9, the attack by azide nucleophile takes place exclusively at the less hindered position, whereas in the case of α -phenyl epoxides such as 3, 6, 7 and 8, the attack takes place predominantly at the benzylic position, although trace amounts of the other regioisomer were detected in the case of epoxides 6, 7 and 8.

In order to compare the regioselectivity of this ring opening reaction, a comparative study was carried out with the classical method. Our findings are shown in Table 2. The high regioselectivity of our catalyst is evident from the yields of the individual regioisomers.

Keywords: Molecular sieves; Azidoalcohol; Regioselectivity; (-)-Cytoxazone.

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Entry	Epoxide	Product	Time (h)	Yield ^a (%)
1			1.0	97
2	O OBn 2	OH N ₃ 2a	1.2	96
3		OH N ₃	1.0	96
4	3 OH OH	3a OH OH	1.3	92
5			1.5	90
6	COOEt	N ₃ COOEt OH	1.5	94 (3)
7	6 MeO 7	6a N ₃ COOEt MeO OH 7a	1.6	95 (2)
8	о он в	N ₃ OH 8a	1.2	95 (2)
9	e e e e e e e e e e e e e e e e e e e		1.2	92

Table 1. Regioselective ring opening of epoxides using NaN₃/CH₃CN/4Å MS

^a Yields of pure isolated product. Yields in the parentheses correspond to the yield of the other regioisomer.

The reason for high regioselectivity exhibited by 4Å molecular sieve in the epoxide opening may be attributed to potassium ion, which is the cation constituent of 4Å molecular sieves.¹¹ Finally we turned our attention towards the possible recycling of the catalyst. The catalyst was filtered off, washed with acetone, dried and reused. In these experiments products were obtained in almost the same yields and regioselectivity.

Seeing the near exclusive formation of the regioisomer 7a (Table 2, entry 4), we decided to employ this method for a synthesis of (–)-cytoxazone 15, a natural product possessing cytokine modulating activity isolated from *Streptomyces* sp. by Osada and co-workers.¹² It is

reported that (–)-cytoxazone exhibits cytokine modulating activity by inhibiting the signalling pathway of Th2 cells¹³ and because of interesting biological activity, this molecule has been the target of various recent synthetic efforts.¹⁴ Sunjic and co-workers synthesized this molecule in the optically pure form using a strategy involving epoxide ring opening with NaN₃. However in their synthesis the yield of the desired regioisomer of the azido alcohol was only 56% and to obtain the final product they employed a kinetic resolution step using *Candida antarctica* lipase (CAL).^{14b}

We started the synthesis of (4R,5R)-cytoxazone from ethyl *p*-methoxycinnamate **10**, which was prepared on

Table 2. Comparative results of regioselectivity

Entry	Epoxide	Product	Conditions ^a	Yield ^b (%)
1	2	2a	А	96
			В	88 (2)
2	3	3a	А	95 (2)
			В	76 (12)
3	6	6a	А	94 (3)
			В	78 (13)
4	7	7a	А	95 (2)
			В	79 (10)
5	8	8a	А	95 (2)
			В	83 (11)

^a A: 1.5equiv NaN₃, 4Å molecular sieves, CH₂Cl₂. B: 2.2equiv NaN₃, 5equiv NH₄Cl, MeOH/H₂O (8:1) reflux.

^b Yields in parentheses are of the other regioisomer.

a large scale in 82% yield by Wittig olefination of *p*-anisaldehyde with triphenyl carbethoxymethyl phosphonium chloride supported on basic alumina under microwave irradiation (Scheme 1).

Instead of resorting to a kinetic resolution step, we decided to construct the epoxide with the desired chirality at this stage. α , β -Unsaturated esters are very good substrates for asymmetric dihydroxylation resulting in excellent enantiomeric excess. Thus when compound **10** when subjected to ADH⁴ using (DHQD)₂PHAL, diol (2*S*,3*R*)-**11** was isolated in 94% yield and 99% ee¹⁵ as a solid, which was treated with *p*-toluenesulfonyl chloride and triethylamine in dichloromethane¹⁶ to give the monotosylated product (2*S*,3*R*)-**12** as a crystalline solid in 95% yield and 98% ee. The next step in the synthesis was the formation of the epoxide (2*R*,3*S*)-**13**, which was effected by treating (2*S*,3*R*)-**12** with K₂CO₃ in methanol¹⁷ whereupon the epoxide (2*R*,3*S*)-**13** was obtained as a gum in 96% yield and 99% ee without epimerization at the C-2 centre, which is a serious problem with several other base/solvent combinations.¹⁸ Azidolysis of this epoxide was performed with NaN₃ and 4Å molecular sieves in acetonitrile to obtain the azido alcohol (2S,3R)-14 in 95% yield and 98% ee. The carbocationic character in the transition state needed for regioselective attack by the nucleophile, at the benzylic position is encouraged by the electron donating effect of the *p*methoxy group.¹⁹ Conversion of (2S,3R)-14 to (-)-cytoxazone was achieved following the procedure described by Nakata and co-workers^{14a} giving (-)-cytoxazone in 48% in overall yield, the spectral data of which were identical to those reported.¹²

In summary we have described a novel procedure for the regioselective ring opening of epoxides, which does not require tedious preparation of the catalyst or harsh conditions like other solid-supported catalysts employed for this purpose. The reusability of this catalyst, high degree of regioselectivity, mild reaction conditions, low cost of the reagent and easy work-up procedure make it an attractive methodology. Further we have described an efficient synthesis of (–)-cytoxazone using this methodology as one of the steps.

Spectral data of some selected compounds: **2a**. IR (chloroform, cm⁻¹): 3420, 2917, 2866, 2101, 1496, 1454, 1279, 1099, 738, 698. ¹H NMR (300 MHz, CDCl₃): $\delta = 7.21$ (5H), 4.43 (s, 2H), 3.81 (m, 1H), 3.45 (d, J = 4Hz, 2H), 2.83 (br s). MS (ESI) m/z = 230.1 (M⁺+Na). Compound **6a**. IR (chloroform, cm⁻¹): 3462, 2983, 2107, 1736, 1736, 1637, 1311, 1256, 1069, 769, 702. ¹H NMR (300 MHz, CDCl₃) $\delta = 7.29$ (5H), 4.85 (d, J = 2.7Hz, 1H), 4.21 (m, 1H), 4.11 (q, J = 4.4Hz, 2H), 1.37 (t, J = 7.1Hz, 3H), 2.11 (br s).



Scheme 1. Reagents and conditions: (a) $(DHQD)_2PHAL$, OsO_4 , *t*-BuOH/H₂O (1:1), 0°C 16h; (b) TsCl, Et₃N, CH₂Cl₂, 5°C, 72h; (c) K₂CO₃, MeOH, rt, 12h; (d) NaN₃, 4Å molecular sieves, CH₃CN, rt, 1.6h; (e) (i) PhCOCl, pyridine, CH₂Cl₂, (ii) PPh₃, THF/H₂O, 50°C, (iii) NaBH₄, MeOH, 0°C (67% in three steps).

MS (ESI) m/z = 258.0 (M⁺+Na). Compound **8a**. IR (chloroform, cm⁻¹): 3408, 2931, 2103, 1443, 1279, 1104, 944, 873, 771. ¹H NMR (300 MHz, CDCl₃) $\delta = 7.21$ (5H), 4.22 (d, J = 6Hz, 1H), 3.59 (m, 1H), 3.28 (d, J = 2.6Hz, 1H), 2.08 (br s, 1H). MS (ESI) m/z = 216.0 (M⁺+Na).

(2*S*,3*R*)-**11**: Solid; mp: 72–73 °C; $[\alpha]_D^{20}$ – 6.1 (*c* 1 methanol); ee: 99%. IR (chloroform, cm⁻¹): 3449, 2982, 2937, 2838, 1735, 1613, 1514, 1465, 1389, 1370, 1302, 1249, 1178, 1031, 834, 771. ¹H NMR (300 MHz, CDCl₃) δ = 7.38 (d, *J* = 8.5 Hz, 2H), 6.84 (d, *J* = 8.5 Hz, 2H), 4.92 (d, *J* = 5.5 Hz, 1H), 4.12 (q, *J* = 4.4 Hz, 2H), 3.85 (s, 3H), 3.25 (d, *J* = 5.5 Hz, 1H), 2.85 (br, 1H), 1.35 (t, *J* = 7.17 Hz, 3H). MS (ESI) *m*/*z* = 263.0 (M⁺+Na). ¹³C NMR (75 MHz, CDCl₃) δ = 13.09, 54.29, 61.11, 73.23, 73.8, 112.8, 124.89, 126.62, 131.07, 158.35, 171.80. Anal. Calcd for C₁₂H₁₆O₅: C, 59.99; H, 6.71. Found: C, 59.80; H, 6.88.

(2*S*,3*R*)-12: Solid; mp 97–98 °C; $[\alpha]_D^{20}$ –46 (*c* 1 methanol); ee: 98%. IR (chloroform, cm⁻¹): 3426, 3025, 2930, 1742, 1614,1514, 1370, 1250, 1176, 1030, 882, 767. ¹H NMR (300 MHz, CDCl₃) δ = 7.57 (d, *J* = 8.2 Hz, 2H), 7.19 (d, *J* = 8.1 Hz, 2H), 7.10 (d, *J* = 8.6 Hz, 2H), 6.71 (d, *J* = 8.6 Hz, 2H), 4.98 (d, *J* = 4.9 Hz, 1H), 4.76 (d, *J* = 5 Hz, 1H), 4.03 (q, *J* = 7 Hz, 2H), 3.75 (s, 3H), 2.82 (br, 1H), 2.41 (s, 3H), 0.98 (t, *J* = 7 Hz, 3H). MS (ESI) *m*/*z* = 417.1 (M⁺+Na). ¹³C NMR (75 MHz, CDCl₃) δ = 12.81, 20.63, 54.05, 60.81, 72.26, 75.59, 76.01, 76.44, 80.57, 95.13, 112.64, 126.65, 127.00, 128.48, 128.55, 131.79, 143.67, 158.57, 165.88.

(2R,3S)-13: Gum; $[\alpha]_{D}^{20}$ +9.1 (*c* 0.8, chloroform); ee: 98%. IR (chloroform, cm⁻¹): 2926, 2936, 2838, 1733, 1612, 1513, 1463, 1301, 1249, 1111, 1030, 833. ¹H NMR (300 MHz, CDCl₃) δ = 7.31 (d, *J* = 8.5 Hz, 2H), 6.88 (d, *J* = 8.5 Hz, 2H), 4.24 (d, *J* = 7 Hz, 1H), 4.02 (q, *J* = 7 Hz, 1H), 3.82 (d, *J* = 4.7 Hz, 1H), 3.70 (s, 3H), 1.30 (t, *J* = 7 Hz, 3H). MS (ESI) *m*/*z* = 222.0 (M⁺). Anal. Calcd for C₁₂H₁₄O₄: C, 64.85; H, 6.35. Found: C, 64.63; H, 6.12.

(2*S*,3*R*)-14: Liquid; $[\alpha]_{20}^{20}$ –23 (*c* 1.0 chloroform); ee: 98%. IR (chloroform, cm⁻¹): 3490, 2965, 2107, 1744, 1612, 1514, 1428, 1252, 1213, 1131, 1028, 827. ¹H NMR (300 MHz, CDCl₃) δ = 7.30 (d, *J* = 8.5 Hz, 2H), 6.88 (d, *J* = 8.5 Hz, 2H), 4.85 (d, *J* = 2.7 Hz, 1H), 4.23 (q, *J* = 7 Hz, 2H), 3.75 (s, 3H), 3.08 (d, *J* = 6.7 Hz, 1H), 1.30 (t, *J* = 7 Hz, 3H). MS (ESI) *m*/*z* = 265.1 (M⁺). ¹³C NMR (75 MHz, CDCl₃) δ = 12.82, 54.60, 56.73, 67.45, 77.51, 77.45, 77.85, 112.81, 125.49, 127.13, 156.43, 168.29. Anal. Calcd for C₁₂H₁₅O₄N₃: C, 54.33; H, 5.70; N, 15.84. Found: C, 54.49; H, 5.83; N, 15.97.

General method for epoxide ring opening: NaN₃ in 2mL CH₃CN were added to powdered 4Å molecular sieves (100 mg) and the mixture stirred for specific time. The reaction mixture was then filtered, the filter pad was washed with CH₃CN and the combined filtrates was evaporated. The crude product was then purified by

chromatography over silica gel using ethylacetate/petroleum ether as eluent.

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- 15. The enantiomeric excess (ee) was measured by HPLC analysis carried out on a Waters 510 HPLC system. Chiracel OD packed in a SS column of 4.6 mm i.d. \times 250 mm dimension was used. Isocratic elution was applied with a mobile phase *n*-hexane 90% and isopropanol 10% at a flow rate 0.8 mL/min, and pressure 125 psi. UV was monitored at 243 nm.
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